# Applications of Likelihood-based inference with non-mechanistic and mechanistic models in infectious disease modeling 

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## Outline

(1) Background
(2) Example 1: Yellow Fever
(3) Example 2: Pandemic Influenza
(4) Example 3: Zika Fever and Guillain-Barré syndrome
(5) Example 4: Dengue Fever
(6) Conclusions

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## Likelihood-based inference

- The likelihood function is the joint probability distribution of observed data expressed as a function of statistical parameters.
- The likelihood function describes a surface over the domain of permissible parameter values.
- The value that is most likely to be the parameter of the joint probability distribution underlying the observed data is the peak of the surface.
- The procedure for obtaining these arguments of the maximum of the likelihood function is known as maximum likelihood estimation.
https://en.wikipedia.org/wiki/Likelihood_function


## Iterated Filtering (lonides, E. L., Breto, C. and King, A. A. (2006))

- Iterated filtering algorithms are a tool for maximum likelihood inference on partially observed dynamical systems.
- Stochastic perturbations to the unknown parameters are used to explore the parameter space. Applying sequential Monte Carlo (the particle filter) to this extended model results in the selection of the parameter values that are more consistent with the data.
- Appropriately constructed procedures, iterating with successively diminished perturbations, converge to the maximum likelihood estimate.
- Iterated filtering methods have so far been used most extensively to study infectious disease transmission dynamics.
https://en.wikipedia.org/wiki/Iterated_filtering


## Iterated Filtering (lonides, E. L., Breto, C. and King, A. A. (2006))

- The data are a time series $y_{1}, \ldots, y_{N}$ collected at times $t_{1}<t_{2}<\cdots<t_{N}$.
- The dynamic system is modeled by a Markov process $X(t)$ which is generated by a function $f(x, s, t, \theta, W)$ in the sense that $X\left(t_{n}\right)=f\left(X\left(t_{n-1}\right), t_{n-1}, t_{n}, \theta, W\right)$ where $\theta$ is a vector of unknown parameters and $W$ is some random quantity that is drawn independently each time $f($.$) is evaluated.$
- An initial condition $X\left(t_{0}\right)$ at some time $t_{0}<t_{1}$ is specified by an initialization function, $X\left(t_{0}\right)=h(\theta)$.
- A measurement density $g\left(y_{n} \mid X_{n}, t_{n}, \theta\right)$ completes the specification of a partially observed Markov process (POMP).
https://en.wikipedia.org/wiki/Iterated_filtering


## Iterated Filtering (lonides, E. L., Breto, C. and King, A. A. (2006))

Input: A partially observed Markov model specified as above; Monte Carlo sample size $J$; number of iterations $M$; cooling parameters $0<a<1$ and $b$; covariance matrix $\Phi$; initial parameter vector $\theta^{(1)}$
for $m=1$ to $M$
draw $\Theta_{F}\left(t_{0}, j\right) \sim \operatorname{Normal}\left(\theta^{(m)}, b a^{m-1} \Phi\right)$ for $j=1, \ldots, J$
set $X_{F}\left(t_{0}, j\right)=h\left(\Theta_{F}\left(t_{0}, j\right)\right)$ for $j=1, \ldots, J$
set $\bar{\theta}\left(t_{0}\right)=\theta^{(m)}$
for $n=1$ to $N$
draw $\Theta_{P}\left(t_{n}, j\right) \sim \operatorname{Normal}\left(\Theta_{F}\left(t_{n-1}, j\right), a^{m-1} \Phi\right)$ for $j=1, \ldots, J$
set $X_{P}\left(t_{n}, j\right)=f\left(X_{F}\left(t_{n-1}, j\right), t_{n-1}, t_{n}, \Theta_{P}\left(t_{n}, j\right), W\right)$ for $j=1, \ldots, J$
set $w(n, j)=g\left(y_{n} \mid X_{P}\left(t_{n}, j\right), t_{n}, \Theta_{P}\left(t_{n}, j\right)\right)$ for $j=1, \ldots, J$
draw $k_{1}, \ldots, k_{J}$ such that $P\left(k_{j}=i\right)=w(n, i) / \sum_{\ell} w(n, \ell)$
set $X_{F}\left(t_{n}, j\right)=X_{P}\left(t_{n}, k_{j}\right)$ and $\Theta_{F}\left(t_{n}, j\right)=\Theta_{P}\left(t_{n}, k_{j}\right)$ for $j=1, \ldots, J$
set $\bar{\theta}_{i}\left(t_{n}\right)$ to the sample mean of $\left\{\Theta_{F, i}\left(t_{n}, j\right), j=1, \ldots, J\right\}$, where the vector $\Theta_{F}$ has components $\left\{\Theta_{F, i}\right\}$
set $V_{i}\left(t_{n}\right)$ to the sample variance of $\left\{\Theta_{P, i}\left(t_{n}, j\right), j=1, \ldots, J\right\}$
set $\theta_{i}^{(m+1)}=\theta_{i}^{(m)}+V_{i}\left(t_{1}\right) \sum_{n=1}^{N} V_{i}^{-1}\left(t_{n}\right)\left(\bar{\theta}_{i}\left(t_{n}\right)-\bar{\theta}_{i}\left(t_{n-1}\right)\right)$

## Plug-and-Play Inference Framework

- The outbreak is modeled as a Partially Observed Markov process (POMP) and makes use of Iterated Filtering and plug-and-play likelihood-based inference frameworks to fit the data.
- Information Criterion quantifying the tradeoff between the goodness-of-fit of a model and its complexity, is employed for model comparison.
- The simulations were conducted deploying the Euler-multinomial integration method with the time-step fixed to be one day.
- The weekly observed cases, $C_{i}$, are assumed to follow a Negative-Binomial (NB) distribution as

$$
\begin{equation*}
C_{i} \sim \mathrm{NB}\left(n=\frac{1}{\tau}, p=\frac{1}{1+\tau Z_{h, i}}\right) \quad \text { with } \quad \text { mean }: \mu_{i}=Z_{h, i} \tag{1}
\end{equation*}
$$

where $\tau$ denotes an over-dispersion parameter that needs to be estimated.

## Information Criterion

The small-sample-size corrected Akaike's Information Criterion (AICc) is a measurement of the trade-off between model complexity and the goodness-of-fit. The AICc is given by:

$$
\begin{equation*}
\mathrm{AICc}=-2 I(\hat{\Theta})+2 k+\frac{2 k(k+1)}{N-k-1} \tag{2}
\end{equation*}
$$

where $N$ is the number of data points and $k$ is the number of free parameters.
Bayesian Information Criterion (BIC) is defined as

$$
\begin{equation*}
\mathrm{BIC}=-2 I(\hat{\Theta})+k \ln N \tag{3}
\end{equation*}
$$

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## Data and Question



Figure: Yellow fever outbreak in Luanda from Dec 5, 2015 to Aug 18, 2016.

## Yellow Fever Model



Figure: Flowchart of the yellow fever model.

## Yellow Fever Model

$$
\begin{align*}
S_{h}^{\prime} & =-a b \frac{I_{v}}{N_{h}} S_{h}-v(t)  \tag{4a}\\
E_{h}^{\prime} & =a b \frac{I_{v}}{N_{h}} S_{h}-\sigma_{h} E_{h}  \tag{4b}\\
A_{h}^{\prime} & =(1-\delta) \cdot \sigma_{h} E_{h}-\gamma_{h} A_{h}  \tag{4c}\\
I_{h}^{\prime} & =\delta \sigma_{h} E_{h}-\gamma_{h} I_{h}  \tag{4d}\\
T_{h}^{\prime} & =\theta \gamma_{h} I_{h}-\kappa_{h} T_{h}  \tag{4e}\\
R_{h}^{\prime} & =v(t)+\gamma_{h} A_{h}+(1-\theta) \cdot \gamma_{h} I_{h}+(1-\eta) \cdot \kappa_{h} T_{h}  \tag{4f}\\
S_{v}^{\prime} & =B_{v}(t)-a c \frac{\psi A_{h}+I_{h}}{N_{h}} S_{v}-\mu_{v} S_{v}  \tag{4~g}\\
E_{v}^{\prime} & =a c \frac{\psi A_{h}+I_{h}}{N_{h}} S_{v}-\sigma_{v} E_{v}-\mu_{v} E_{v}  \tag{4h}\\
I_{v}^{\prime} & =\sigma_{v} E_{v}-\mu_{v} I_{v} \tag{4i}
\end{align*}
$$

## Yellow Fever Model (non-mechanistic $m(t)$ )

$$
\begin{equation*}
\text { weekly deaths: } Y_{h, i}=\int_{\text {week } i} \eta \kappa_{h} T_{h} d t \tag{5a}
\end{equation*}
$$ weekly reported cases: $Z_{h, i}=\int_{\text {week } i}[\theta+\rho \cdot(1-\theta)] \cdot \gamma_{h} I_{h} d t$

$$
\begin{align*}
& N_{h}=S_{h}+E_{h}+A_{h}+I_{h}+T_{h}+R_{h}+\sum_{i} Y_{h, i}  \tag{6a}\\
& N_{v}=S_{v}+E_{v}+I_{v} \tag{6b}
\end{align*}
$$

$$
\begin{equation*}
N_{v}(t)=m(t) \cdot N_{h} \tag{7}
\end{equation*}
$$

where $m(t)$ is modeled as an exponential cubic spline function.

## Vaccination Scenarios

A simple way to estimate the recovered (including vaccinated) population

$$
\begin{equation*}
R_{h}(t)=V\left(t-t_{0}\right) \cdot N_{h}+\varepsilon_{0}+\varepsilon(t) \tag{8}
\end{equation*}
$$

where $V(t)$ is the vaccination coverage, $\varepsilon_{0}$ denotes the prior immunity and $\varepsilon(t)$ denotes the cumulative infected cases up to time $t$.

- Baseline scenario: actual vaccination campaign as experienced in Luanda;
- Alternative scenarios: 60, 120 and 180 days ( $\equiv$ no vaccination) delays of vaccination campaign

The total reported cases as well as total deaths will be evaluated by the model for each of the different vaccination scenarios.

## Basic Reproduction Number

$$
\begin{equation*}
\mathcal{R}_{0}=\sqrt{[\psi \cdot(1-\delta)+\delta] \cdot \frac{a^{2} b c m}{\gamma_{h}} \cdot \frac{\sigma_{v}}{\mu_{v}\left(\sigma_{v}+\mu_{v}\right)}} \tag{9}
\end{equation*}
$$

or

$$
\begin{equation*}
\mathcal{R}_{0}=[\psi \cdot(1-\delta)+\delta] \cdot \frac{a^{2} b c m}{\gamma_{h}} \cdot \frac{\sigma_{v}}{\mu_{v}\left(\sigma_{v}+\mu_{v}\right)} \tag{10}
\end{equation*}
$$

Asymptomatic Infectivity Scenarios
1 : $85 \%$ asymptomatic $(\delta=15 \%)$ and weak infectivity $(\psi=0.1)$
2 : $85 \%$ asymptomatic $(\delta=15 \%)$ and strong infectivity $(\psi=0.5)$

## Model Fitting



Figure: Fitting results under two scenarios of asymptomatic infectivity status.

## Impact of Vaccination Delays






Figure: Simulation results of asymptomatic-1 scenario under three deferred vaccination campaign scenarios: 60-day delay in panels (a,b), 120-day delay in panels ( $\mathrm{c}, \mathrm{d}$ ) and 180-day delay in panels (e,f).

Table: Impacts of vaccination campaign delay under asymptomatic 1 scenario.

| Scenario | Total reported cases | Total deaths |
| :---: | :---: | :---: |
| Observed | 941 | 73 |
| Baseline model | $1026[540,1797]$ | $77[35,139]$ |
| 60 days delay | $3143[1604,5584]$ | $233[119,411]$ |
| 120 days delay | $5450[2751,9611]$ | $400[203,724]$ |
| 180 days delay | $6242[3139,10919]$ | $444[226,787]$ |

"* $[*, *]$ " denotes the simulation median with $95 \%$ Confidence Interval (C.I.).

## Human behavioral model (mechanistic)

$$
\begin{equation*}
m(t)=m_{\text {base }}+k \cdot \exp \left[-D_{h}\left(t-t_{\text {lag }}\right)\right] \tag{11}
\end{equation*}
$$

Here $m_{\text {base }}$ is a constant term, $k$ is a parameter controlling the strength of the death-induced human reaction, $D_{h}(t)$ is the yellow fever deaths of time $t$ and $t_{\text {lag }}$ is the lag time for the population reacting to the yellow fever death situation. We use linear interpolation to convert the weekly death into a continuous time function. The fitting results for this simple human behavior model are shown in the following figure, with $t_{\text {lag }}=1$ week fixed.

## Human behavioral model



Figure: Fitting results under consideration of humans behavior (with one-week time lag, $t_{\text {lag }}=1$ ).

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## 1918 Influenza in London, England (non-mechanistic model)

Fig. 2 Excess P\&I mortality, realizations of the best model and likelihood profiles for four parameters. The top panel a shows the observed excess P\&I mortality (blue, as in Fig. 1), together with box plots of 1,000 realizations of the best model. Panel b shows the AICc as a function of $N_{\mathrm{B}}$, the number of $B$-spline basis functions. Panels c-e show likelihood profiles for $1 / \gamma, \phi$ and $\mathcal{R}_{0}$ (with horizontal lines that define the $95 \%$ confidence intervals; see "Methods")






## 1918 Influenza in England (mechanistic model)



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## Zika fever in Brazil (temperature-driven)



## Zika fever in Brazil (non-mechanistic)



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## Dengue fever in Kaohsiung and Tainan



## Dengue Fever in Kaohsiung and Tainan



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## Conclusions

- Plug-and-play likelihood-based inference framework is a powerful tool for disease modeling.
- While mechanistic models require additional assumptions and knowledge, non-mechanistic models do not.
- The comparison between fitting mechanistic models and fitting non-mechanistic models to data helps gain insight on possible mechanisms underlying disease outbreaks.


## THANK YOU!

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